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Synthesis of L- α -phosphatidyl-D-*myo*-inositol 3,5-bisphosphate from D-glucose

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Abstract—Efficient synthesis of L- α -phosphatidyl-D-*myo*-inositol 3,5-bisphosphate was achieved from 1,2,5,6-diisopropylidene-D-glucose by utilizing ring-closing metathesis and catalytic OsO₄ dihydroxylation. © 2001 Elsevier Science Ltd. All rights reserved.

L- α -Phosphatidyl-D-*myo*-inositol (PI) and its phosphates (PIPn) play important roles as second messengers in intracellular signal transduction.¹ We have already reported the synthesis of various lipid analogs of PIPns as artificial second messengers and biochemical probes.^{2–4} Recently, L- α -phosphatidyl-D-*myo*-inositol 3,5-bisphosphate (PI 3,5-P2, **1a**) was identified as a novel phosphoinositide (Fig. 1).⁵ In mouse cells, PI 3,5-P2 is synthesized by phosphorylation of PI 3-P at the D-5 position by Fab 1.⁶ Several studies have shown that PI 3,5-P2 production is essential for sorting membrane proteins into the lumen of the yeast vacuole⁷ and for the maintenance of vacuolar size.⁸ Although PI 3,5-P2 may be important for membrane homeostasis, its biological role has not yet been defined clearly.

To supply suitably protected optically active *myo*-inositol derivatives with appropriate stereogenic centers, Ferrier rearrangement,^{9,10} pinacol coupling¹¹ and ringclosing metathesis (RCM)¹² of chiral intermediates derived from precursors such as carbohydrates have been performed in addition to optical resolution of *myo*-inositol derivatives. In 1996, we reported the facile asymmetric synthesis of *myo*-inositol derivatives via a 1,7-diene intermediate, which was synthesized from tartaric acid with complete stereocontrol.^{11d} Since then, several reports have appeared by other groups employing 1,7-dienes as key synthetic intermediates.^{11e,12,13} In this communication, we describe an efficient asymmetric synthesis of PI 3,5P2 (**1b**), a novel lipid analog of **1a**,^{14,†} by RCM of a properly protected 1,7-diene (**4**)



Figure 1.

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^{\dagger} According to our previous results, PI and PIP3 lipid analogs having a short saturated side chain at the *sn*-2 position were excellent substrates of PI 3-kinase and PIP3 5-phosphatase, respectively, while analogs with a long saturated side chain were not. Therefore, we employed butyrate as the *sn*-2 side chain of PI 3,5-P2, taking account of the potential biochemical application. See Ref. 3.



Scheme 2. (a) NaH, BnBr, DMF, 89%; (b) 60% aq. AcOH, rt, 95%; (c) NaIO₄, aq. NaHCO₃, CH₂Cl₂, rt, 2 h, 73%; (d) vinylmagnesium bromide (3.0 equiv.), CuBr–Me₂S (3.4 equiv.), THF–Me₂S, $-40^{\circ}C \rightarrow rt$, 2 h, 59%; (e) NaH, BnBr, DMF, 95%; (f) 80% AcOH, 70°C, 2 days, 92%; (g) MePPh₃Br, *n*-BuLi, ether, rt, overnight, 68%; (h) Cl₂(PCy₃)₂Ru=CHPh (10 mol%), CH₂Cl₂, rt, 1 h, 85%; (i) NaH, PMBCl, DMF, 92%; (j) OsO₄ (5 mol%), quinuclidine (5 mol%), NMO, CH₂Cl₂, rt, overnight, 98% (1:1 mixture of diastereoisomers); (k) BzCl (1.2 equiv.), pyridine, DMAP, 67% (+dibenzoates 27%).

derived from D-glucose, followed by catalytic OsO_4 dihydroxylation (Scheme 1).

The 1,2-addition of an organocopper reagent prepared from vinylmagnesium bromide and CuBr–Me₂S to the known aldehyde (6) prepared from 1,2,5,6-diisopropylidene-D-glucose gave the desired allyl alcohol (7) as the sole product (Scheme 2).^{15,16,‡} After protection of

the C-2 hydroxy group as the benzyl ether, isopropylidene acetal was hydrolyzed with aq. AcOH to give the hemiacetal (8), which was subjected to Wittig methylenation to give the 1,7-diene (9). Ring-closing metathesis of 9 using Grubbs' catalyst $Cl_2(PCy_3)_2Ru=CHPh^{17}$ gave the cyclohexene (10) in 85% yield, and then two hydroxyl groups were protected as the PMB ether. The dihydroxylation of 11 utilizing a catalytic amount of osmium tetroxide with quinuclidine and NMO as co-oxidant gave the *cis*-diols (12a, 12b) in 98% yield as an equimolar mixture of diastereoisomers by ¹H NMR analysis. Selective benzoylation of equatorially oriented hydroxyl groups of this mixture gave the benzoates (13a, 13b), which were separated by silica gel column chromatography. Benzyloxymethylation of the desired 13a followed by methanolysis of the benzoate afforded the appropriately protected *myo*-inositol (15) (Scheme 3).

The coupling of 15 with phosphoramidite (16), in the presence of 1H-tetrazole, followed by m-CPBA oxida-

[‡] The 1,2-addition of vinylmagnesium bromide to the aldehyde **6** took place with no selectivity (7:*epi*-7=1.25:1). Stereochemistry of these products was established by the comparison of the ¹H NMR spectrum of **11** derived from a known bisallyl alcohol. See Ref. 11d. ¹H NMR of **11** (CDCl₃, 500 MHz) δ 3.68 (1H, d, *J*=5.1 Hz), 3.69 (1H, d, *J*=5.1 Hz), 3.79 (6H, s), 4.17 (1H, d *J*=5.1 Hz), 4.18 (1H, d, *J*=5.1 Hz), 4.60 (1H, d, *J*=11.1 Hz), 4.64 (1H, d, *J*=11.1 Hz), 4.67 (1H, d, *J*=11.6 Hz), 4.71 (1H, d, *J*=11.6 Hz), 4.77 (1H, d, *J*=10.4 Hz), 4.84 (1H, d, *J*=10.4 Hz), 4.85 (1H, d, *J*=11.0 Hz), 4.92 (1H, d, *J*=11.0 Hz), 5.69 (1H, s), 5.70 (1H, s), 6.82 (2H, d, *J*=8.6 Hz), 6.84 (2H, d, *J*=8.6 Hz), 7.22–7.26 (4H, m), 7.28–7.37 (10H, m)



Scheme 3. (a) BOMCl, diisopropylethylamine, tetra-*n*-butylammonium bromide, CH_2Cl_2 , 70°C, 2 days, 30% (recovered 13a 68%); (b) K₂CO₃, MeOH, rt, 4 h, quant.



Scheme 4. (a) 15, 1*H*-tetrazole (8.0 equiv.), CH₂Cl₂, rt, 2.5 h, then *m*-CPBA (4.2 equiv.), $-78^{\circ}C \rightarrow rt$, 45 min, quant.; (b) DDQ (3.0 equiv.), wet CH₂Cl₂, rt, 2.5 h, 52%; (c) (BnO)₂PNEt₂ (5.7 equiv.), 1*H*-tetrazole (6.0 equiv.), CH₂Cl₂, rt, 2.5 h, then *m*-CPBA (8.0 equiv.), $-78^{\circ}C \rightarrow rt$, 30 min, 82%; (d) H₂ (4.3 kgf/cm²), Pd black, NaHCO₃, *tert*-BuOH/H₂O, quant.

tion gave the phosphodiester 17 (Scheme 4). PMB protecting groups of 17 were oxidatively removed by 2,3-dichloro-5,6-dicyano-1,4-benzoquinone (DDQ) and the free hydroxyl groups were phosphorylated by the amidite method to give the fully protected PI 3,5-P2 precursor. Finally, hydrogenolysis with Pd black as a catalyst in the presence of NaHCO₃ gave PI 3,5-P2 (1b) as the sodium salt.[§]

In conclusion, we achieved efficient construction of the key 1,7-diene intermediate, which is adaptable for the synthesis of not only a series of inositol analogs, but also various six-membered cyclitols such as condritol B, and we also successfully completed the total synthesis of PI 3,5-P2 utilizing RCM and catalytic OsO_4 dihydroxylation. Clarification of the biochemical function of PI 3,5-P2 as a second messenger using this synthetic product as a biological tool is in progress.

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[§] Spectral data for **1b** (Na salt): ³¹P NMR (D₂O, 202 MHz, 5% K₃PO₄ external reference): δ –2.87, –4.16, –5.46; FABMS m/z (*m*-NBA+Gly+NaCl): 875 (M–H+2Na)⁺, 897 (M–2H+3Na)⁺, 919 (M–3H+4Na)⁺, 941 (M–4H+5Na)⁺, 963 (M–5H+6Na)⁺; HRMS (FAB) m/z: calcd for C₃₁H₅₈O₁₉P₃Na₄ (M–3H+4Na)⁺, 919.2376, found: 919.2425.

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